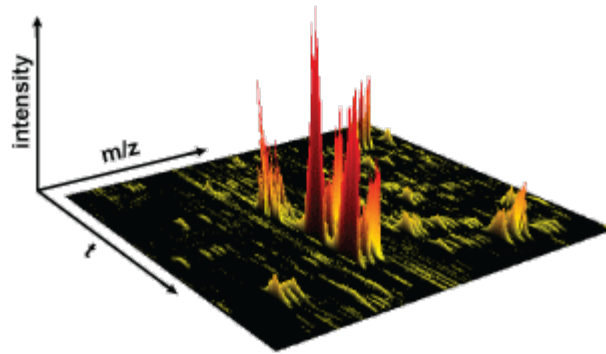


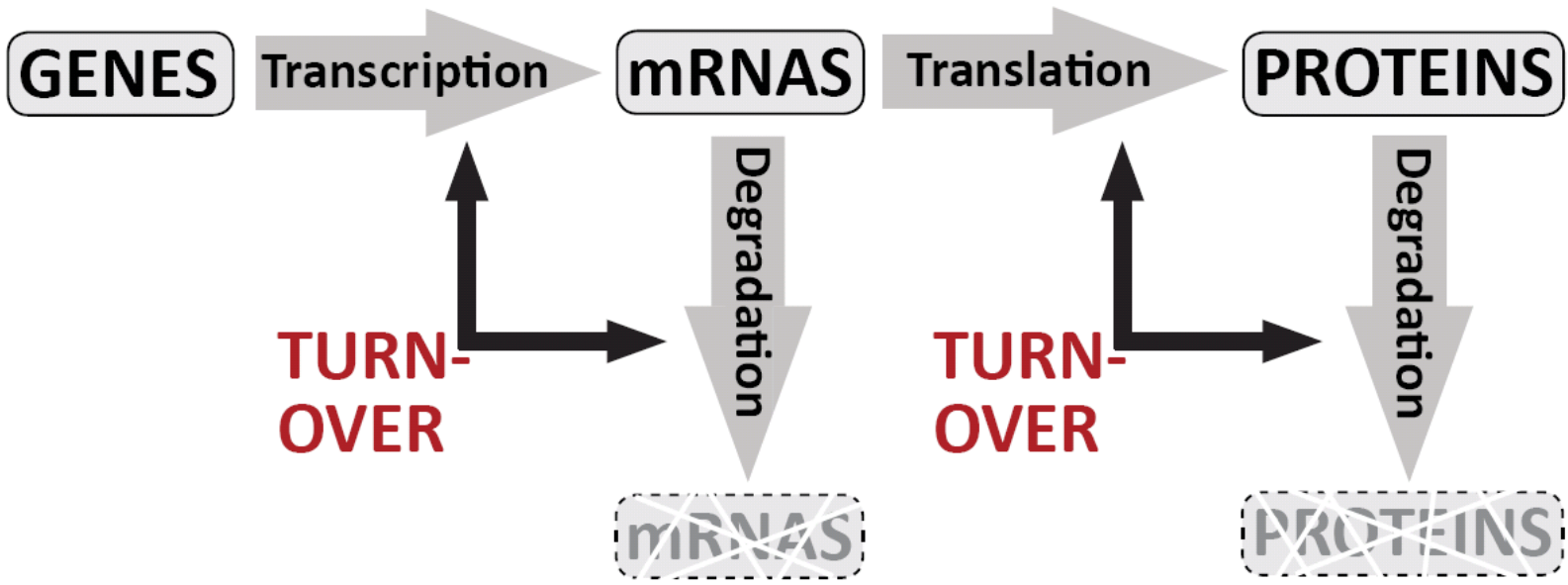
# Genome-wide analysis of protein and mRNA half-lives reveals dynamic properties of mammalian gene expression

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[www.mdc-berlin.de/selbach](http://www.mdc-berlin.de/selbach)

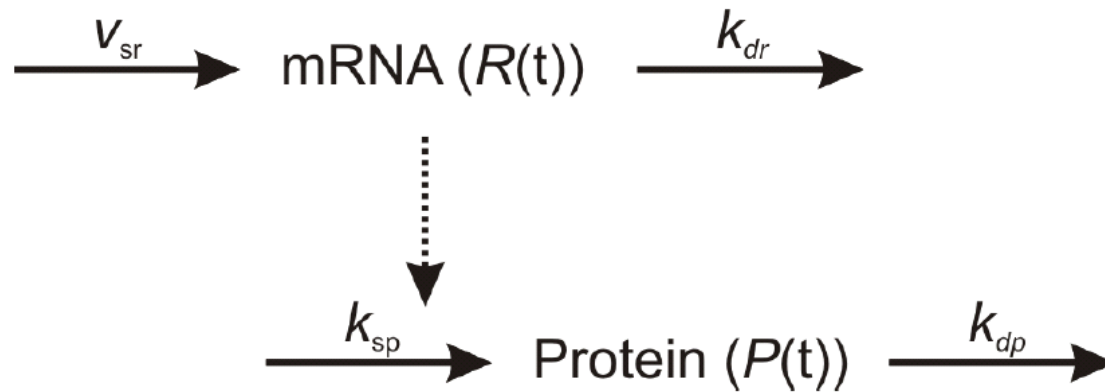
# Dynamic regulation of gene expression



➤ Dynamic properties of gene expression depend on protein and mRNA levels and half-lives

# Mathematical modeling of gene expression

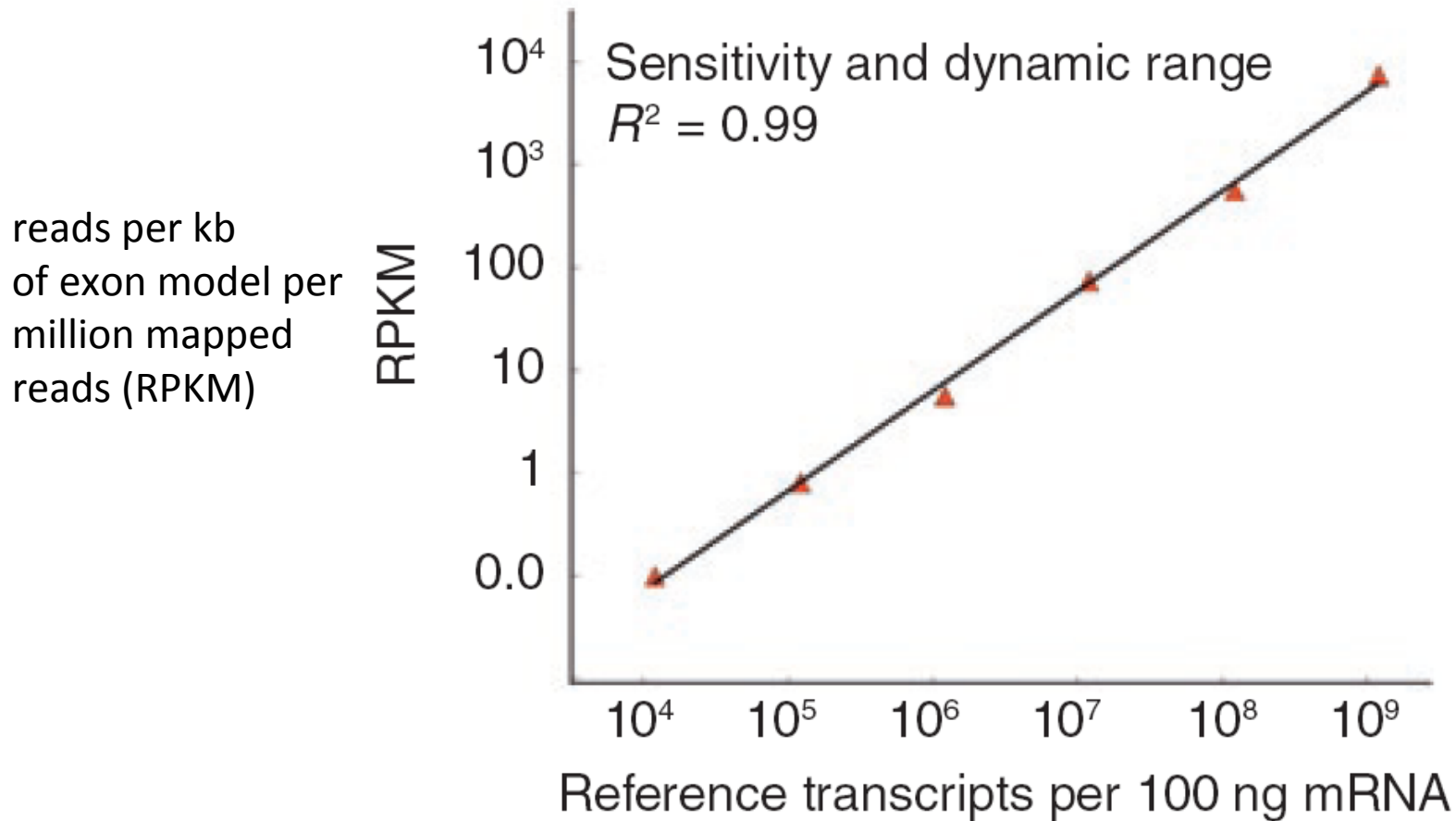
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$$\frac{dR}{dt} = v_{sr} - k_{dr}R$$

$$\frac{dP}{dt} = k_{sp}R - k_{dp}P$$

# Absolute mRNA levels: next generation sequencing



# Methods for absolute protein quantification

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## Stable isotope-based methods:

- AQUA peptides
- iTRAQ
- QconCAT
- absolute SILAC

Gerber et al., PNAS, 2003

Ross et al., MCP, 2004

Beynon et al., N. Meth., 2005

Hanke et al., JPR, 2008

## Label free / computational methods:

- protein abundance index (PAI/emPAI)
- normalized spectral abundance factor (NSAF)
- absolute protein expression measurements (APEX)
- top3 peptide intensities (top3)

Ishihama et al, MCP, 2005

Mosley et al., J.Prot., 2009

Lu et al., Nat. Biotech, 2007

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- protein abundance index (PAI/emPAI)
- normalized spectral abundance factor (NSAF)
- absolute protein expression measurements (APEX)
- top3 peptide intensities (top3)
- intensity-based absolute quantification (iBAQ)

Ishihama et al, MCP, 2005  
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Schwanhaeusser et al., unpublished

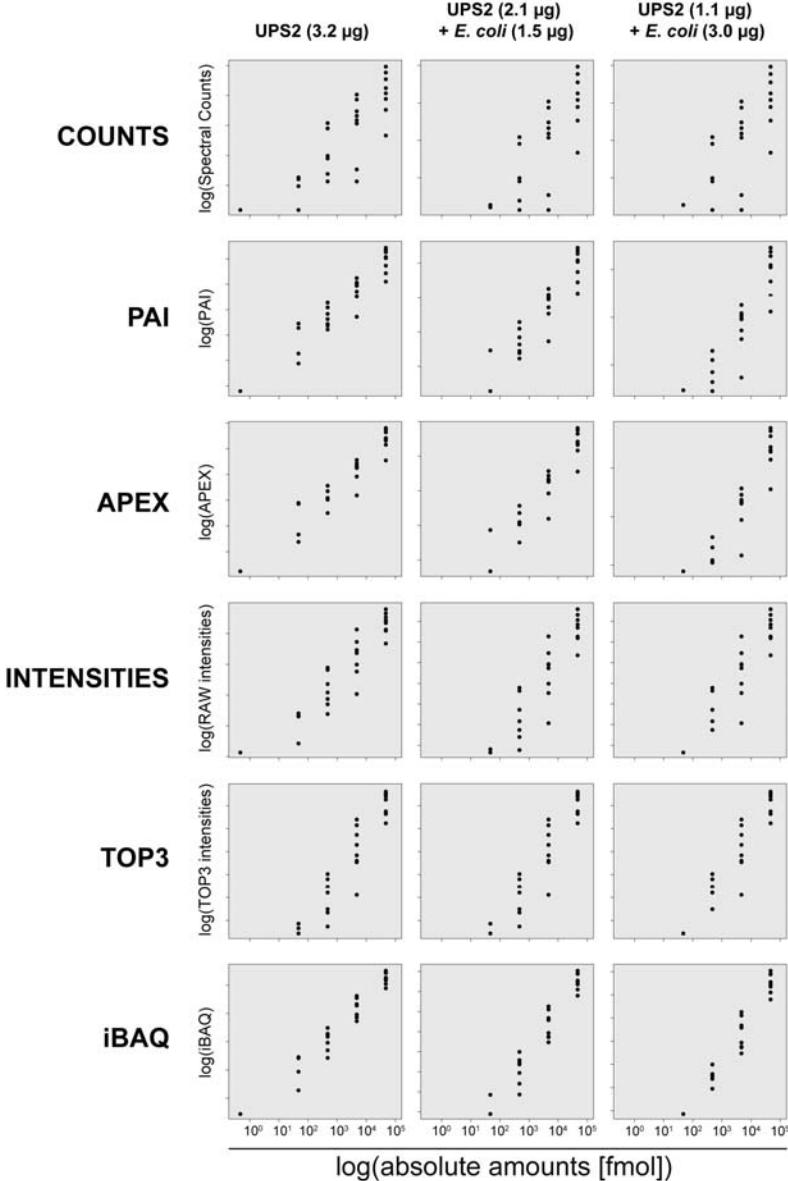
# Benchmarking different methods

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- should be independent of protein sequence
- be accurate over several orders of magnitude
- should not be affected by overall sample complexity

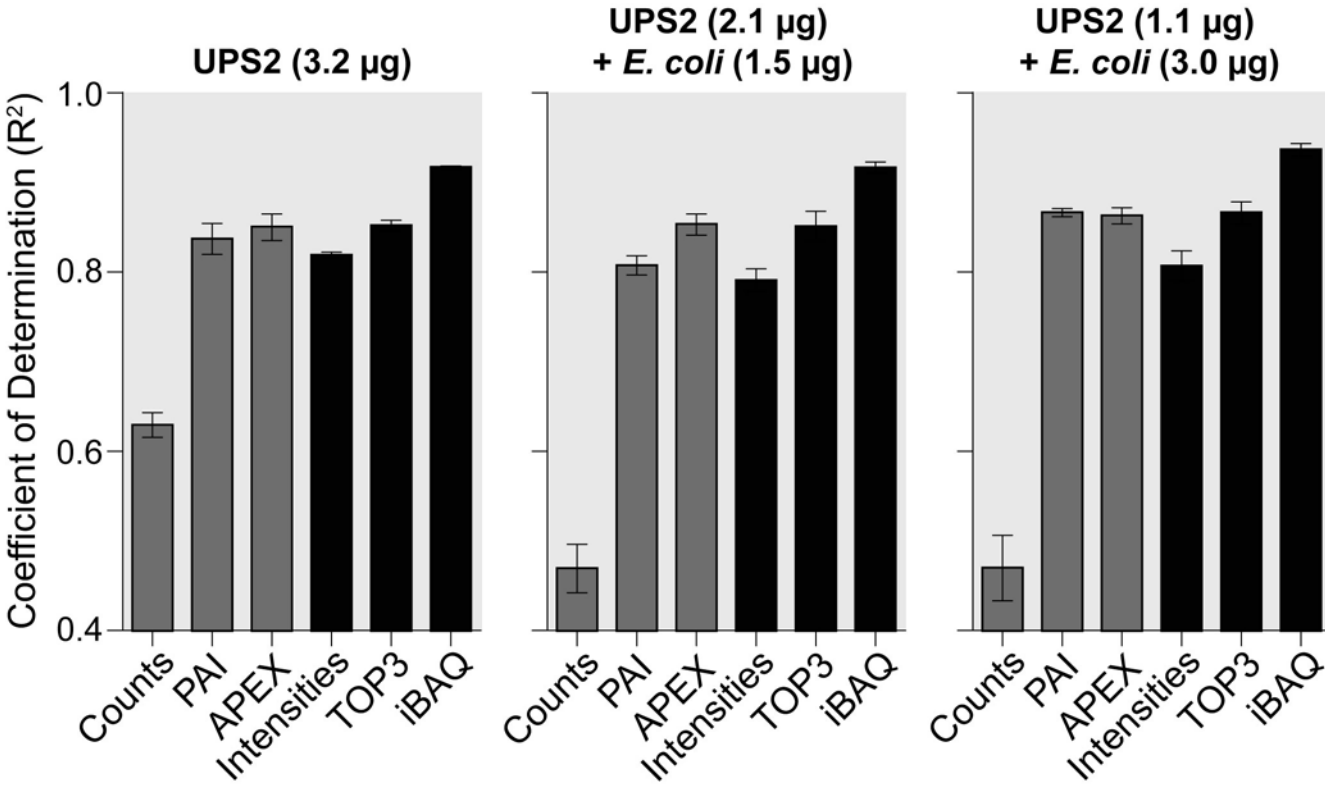
48 recombinant  
human proteins over  
six orders of magnitude  
(UPS2, Sigma)      +      *E. coli* lysate  
(different amounts)

# Benchmarking different methods

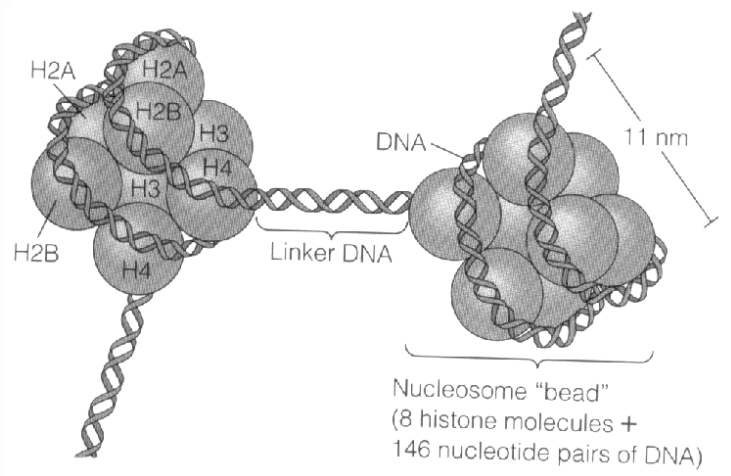
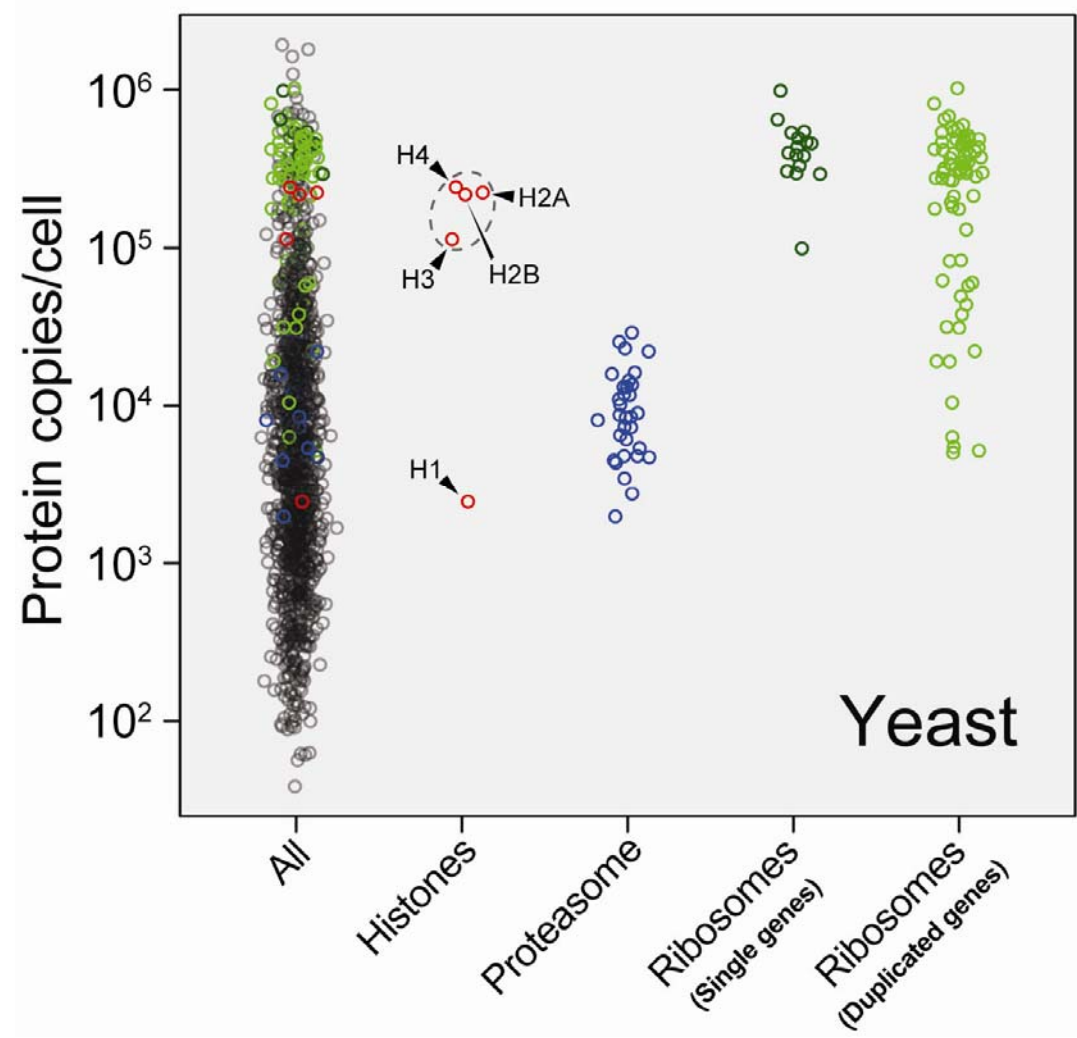




# Benchmarking different methods



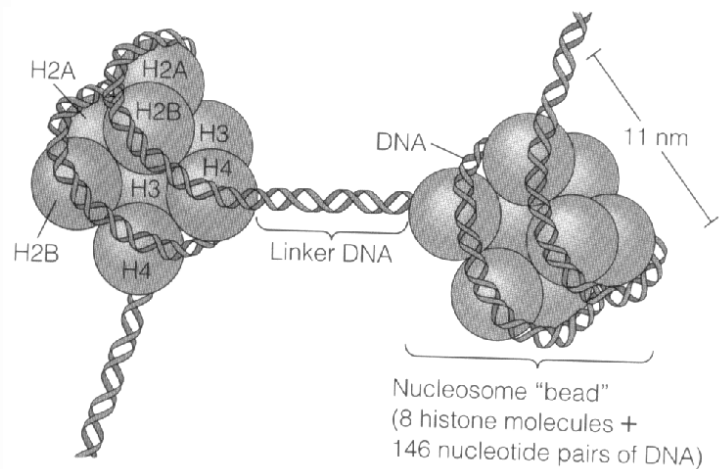
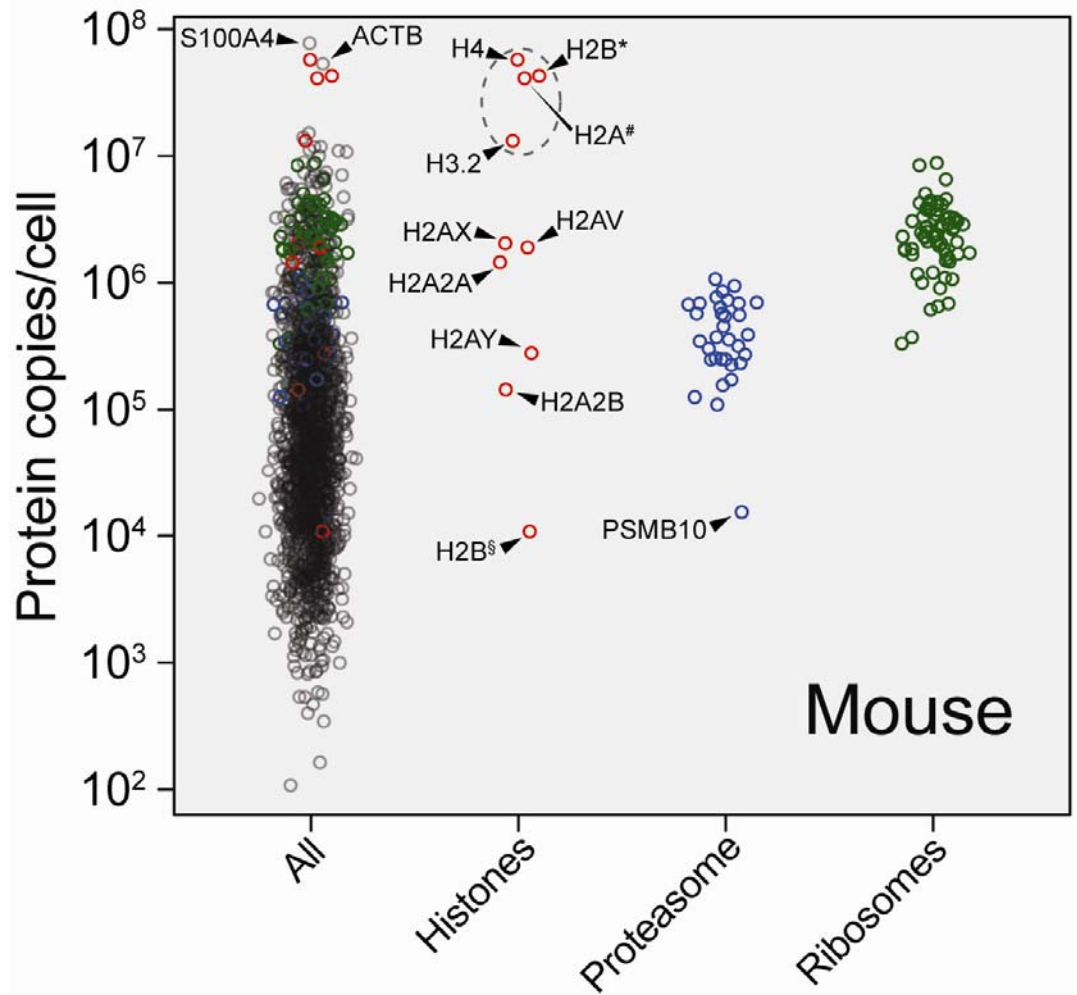
# Cellular copy numbers of yeast and mouse (NIH3T3) proteins



Yeast nucleosomes:  
 12 Mb genome / 150 bp  
 = 80,000 nucleosomes

iBAQ: ~100,000  
 nucleosomes

# Cellular copy numbers of yeast and mouse (NIH3T3) proteins



mouse nucleosomes:  
 2.6 Gb haploid genome size x 2  
 / 150b = 3.5 x 10<sup>7</sup> nucleosomes

iBAQ: ~ 3 x 10<sup>7</sup> nucleosomes

# Summary I: intensity-based absolute quantification (iBAQ)

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- most accurate estimate of absolute protein abundance
- all available information is used (intensities of all peptides)
- spiked-in standard proteins minimize variability in sample handling
- very simple approach:
  - universal standards can be used (AQUA, absolute SILAC)
  - no complex machine learning procedures required (APEX)
  - no need for three different MS methods (top3)

# Large scale analysis of mRNA and protein turnover

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Classical approach:

- inhibition of transcription / translation using drugs
- track mRNA / protein decay over time

Problem:

- drug treatment has severe side effects and ultimately kills the cells under study

# Labeling with isotopes: Rudolf Schoenheimer

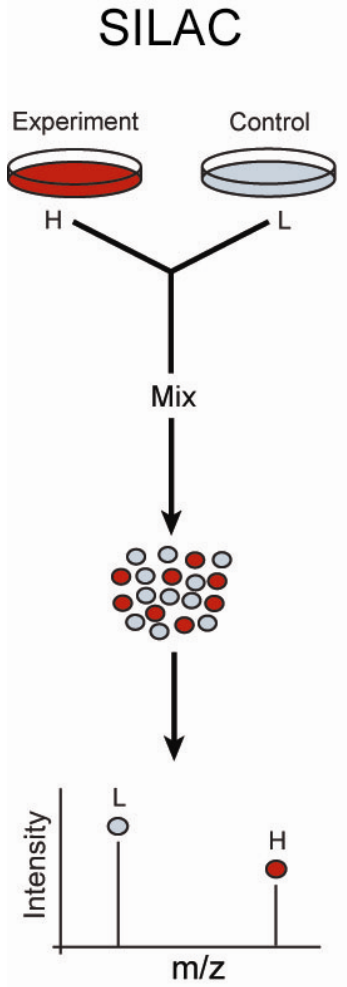


**Deuterium as an Indicator in the Study of Intermediary Metabolism, Schoenheimer and Rittenberg, J. Biol. Chem., 1935**

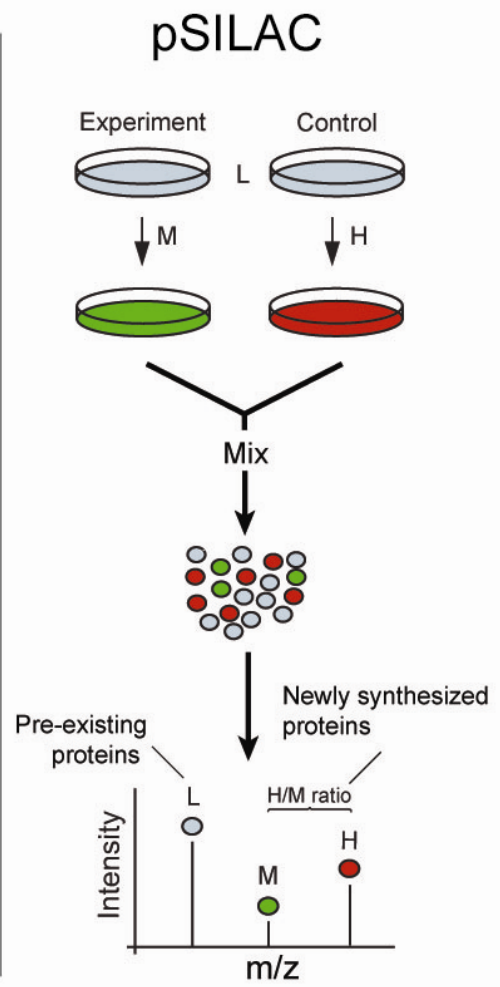
*„In order successfully to label a physiological substance, it is essential that the chemical and physical properties of the labeled substance be so similar to the unlabeled one that the animal organism will not be able to differentiate between them. The chemist, on the other hand, must be able to distinguish and to estimate them in small quantities and at high dilutions. A possibility for such a label is the use of an isotope.”*

Rudolf Schoenheimer, 1898-1941

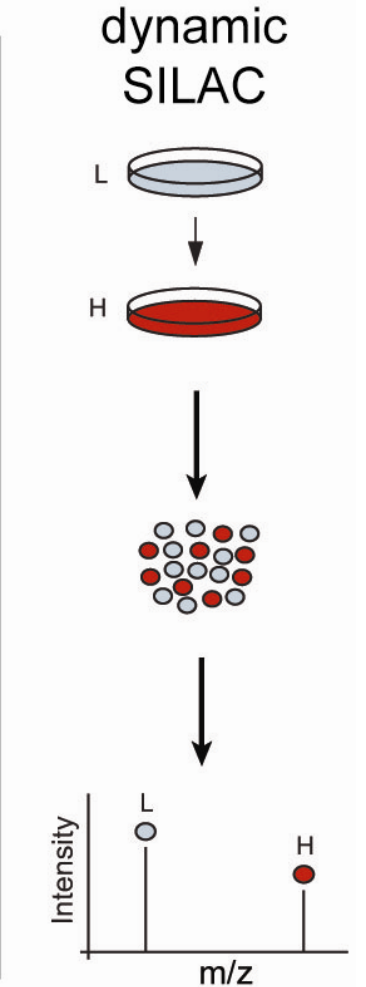
# Measuring protein dynamics using SILAC



Relative changes in protein levels  
(Ong et al., 2002)

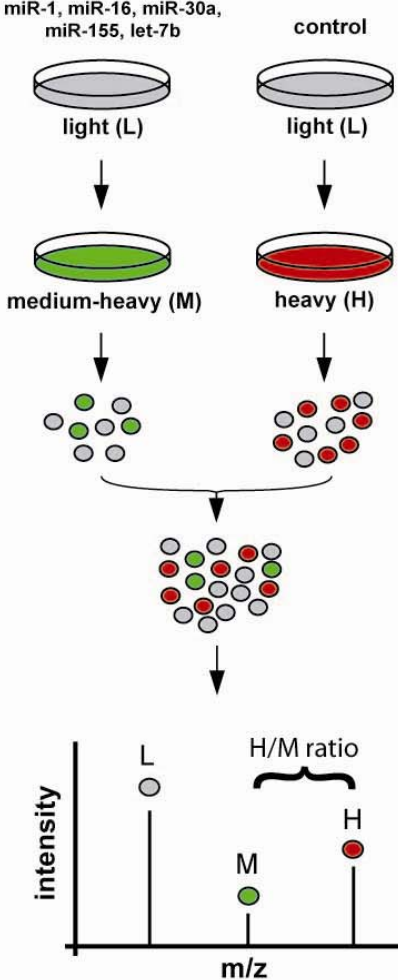
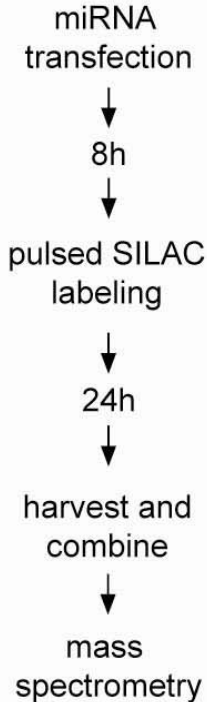


Relative changes in protein production  
(Schwanhaeusser et al, 2009)



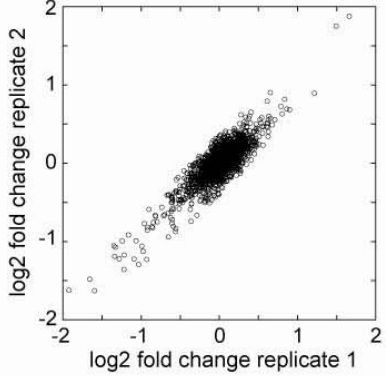
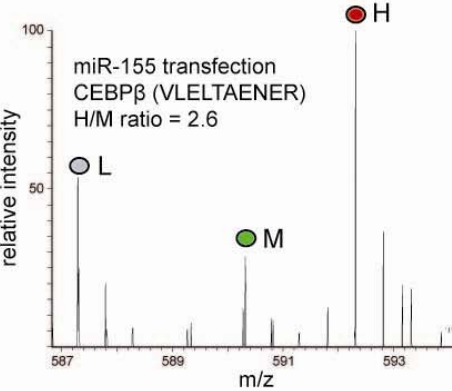
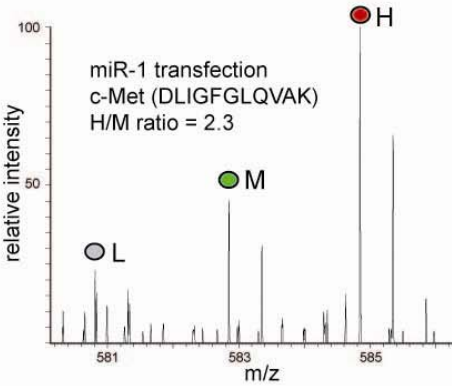
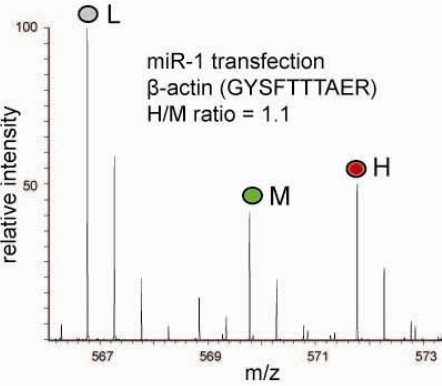
protein turnover  
(Doherty et al., 2009)

# pSILAC identification of miRNA targets



proteins:

- produced in M
- produced in H
- preexisting





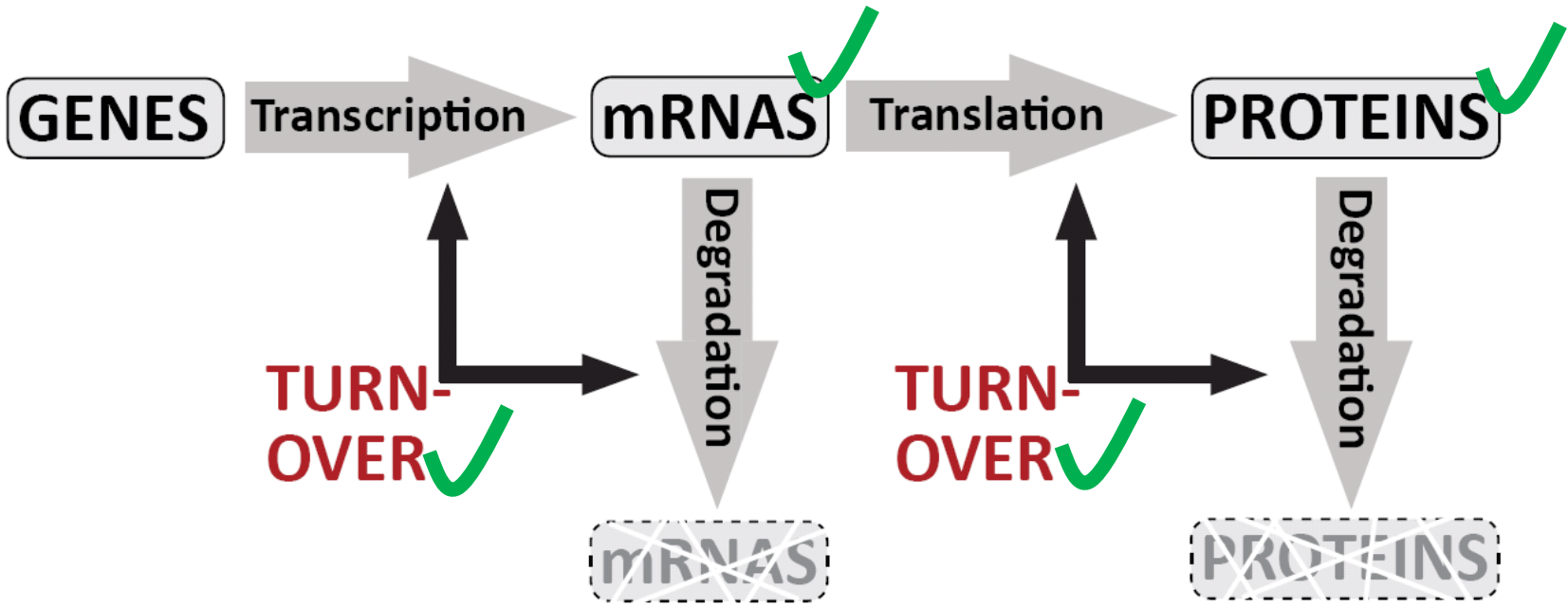
# Metabolic pulse labeling of proteins and mRNAs

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**High-resolution gene expression profiling for simultaneous kinetic parameter analysis of RNA synthesis and decay.** RNA. 2008 Sep;14(9):1959-72.

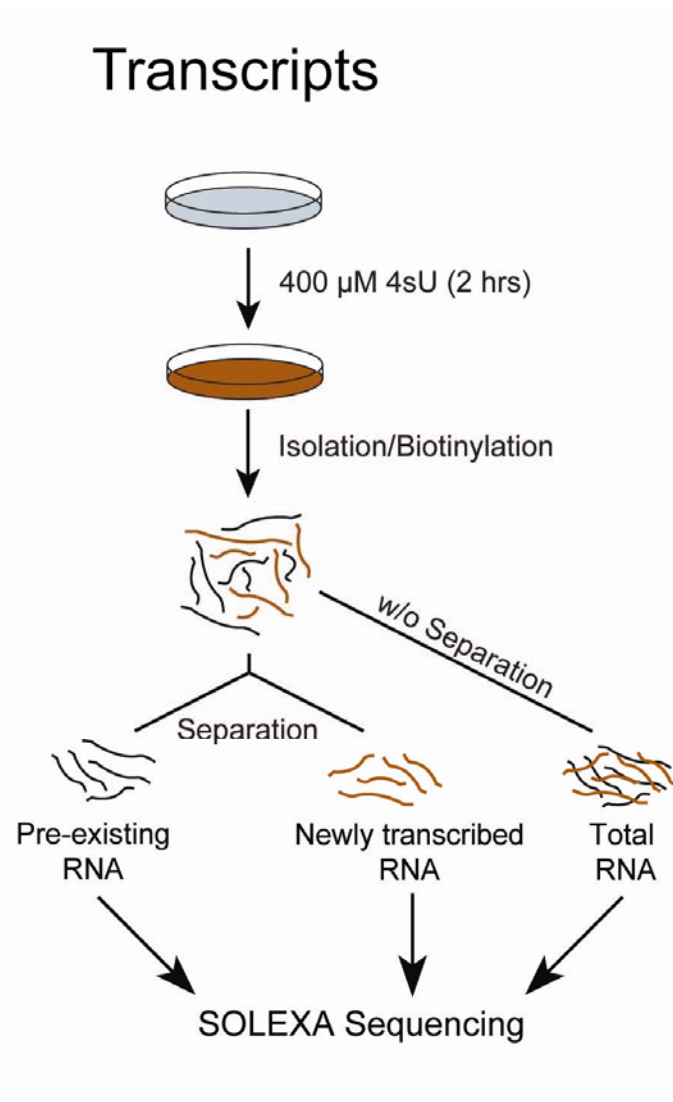
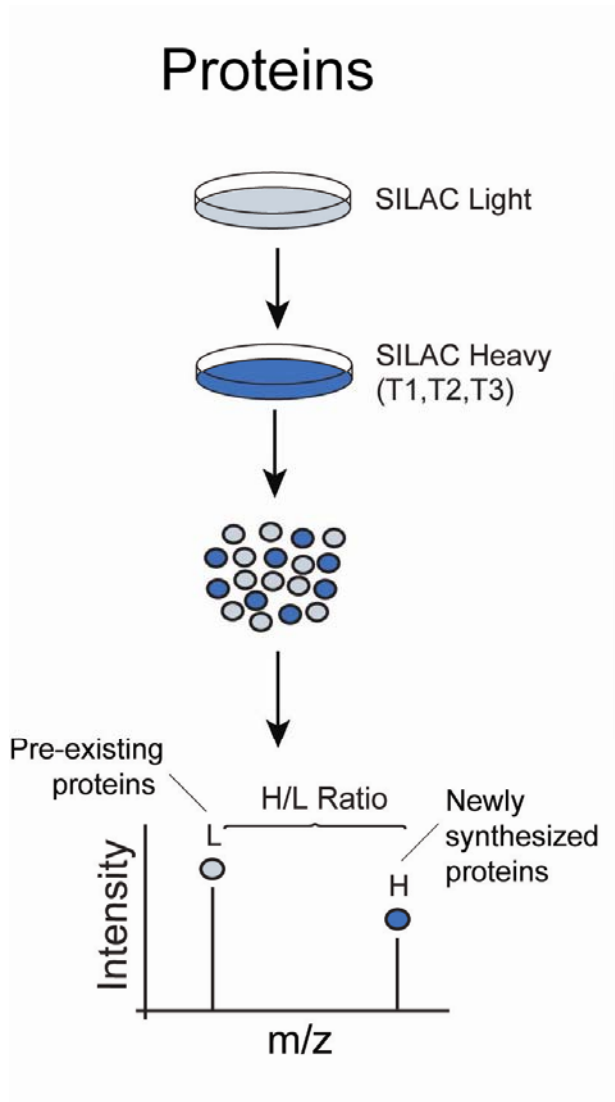
Dölken L, Ruzsics Z, Rädle B, Friedel CC, Zimmer R, Mages J, Hoffmann R, Dickinson P, Forster T, Ghazal P, Koszinowski UH. Max von Pettenkofer-Institute, Ludwig Maximilians-University Munich, Munich 80337, Germany.  
doelken@mvp.uni-muenchen.de

# Dynamic regulation of gene expression

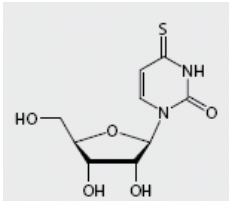


➤ Dynamic properties of gene expression depend on protein and mRNA levels and half-lives

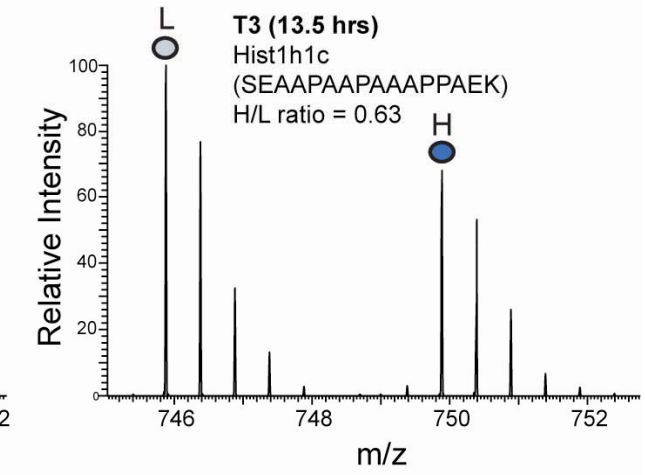
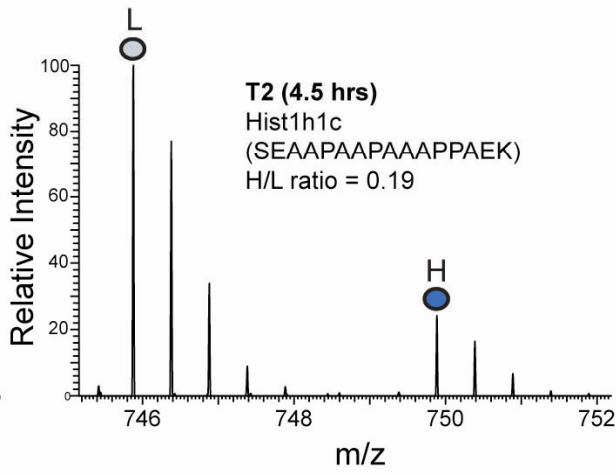
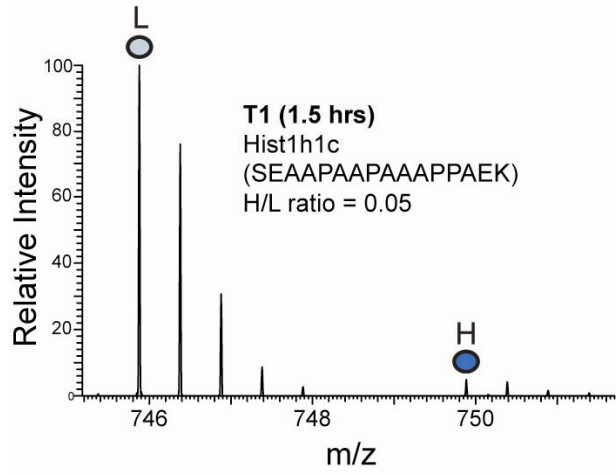
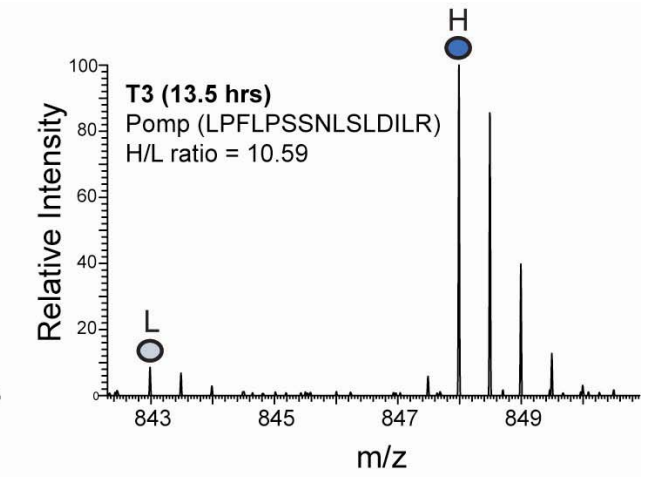
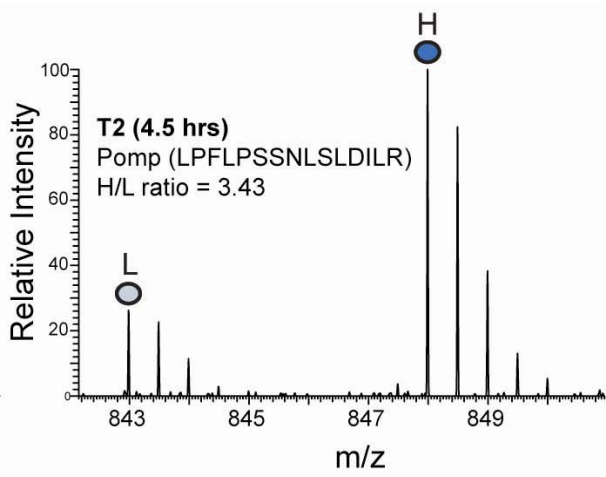
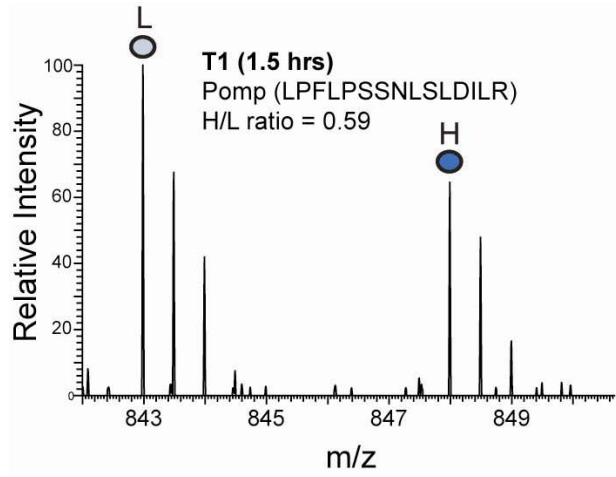
# Dynamic properties of mammalian gene expression: mRNA and protein levels and half-lives



### 4-Thiouridine



# Metabolic pulse labeling of proteins



# Summary

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- parallel metabolic pulse labeling with SILAC and 4SU can be used to assess protein and mRNA turnover at the same time (unperturbed system, no inhibitors)
- deep sequencing and mass spectrometry can quantify protein and mRNA half-lives and absolute cellular protein and mRNA copy numbers (iBAQ and RPKM)
- while mRNA and protein levels are correlated, half-lives are very different
- important consequences for dynamic properties of gene expression
  - energy constraints
  - posttranscriptional regulation (like RNA-binding proteins)
  - dynamic behavior

# Thank you!



Christian Sommer

Dorothea Busse  
Jana Wolf

Wei Chen  
Na Li

Björn Schwanhäusser

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